



Stephanospora mayana (Stephanosporaceae, Russulales), a new sequestrate fungus from Yucatán Peninsula, Mexico

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Abstract

Stephanospora mayana is presented as a new species from the Yucatán Peninsula, Mexico. This species is distinguished by the yellowish pileus, basidiospores with a small corona ($4-6 \times 1-2.5 \mu m$), and variable size ($8.0-17.0 \times 6.0-11.0$), thin pileus ($21-40 \mu m$) and the ecological association to lowland forest with Haematoxylum campechianum, Gymnopodium floribundum, Coccoloba diversifolia, Metopium brownei and Pinus caribaea. It differs from the American species of Stephanospora, like S. michoacanensis and S. chilensis, by its larger basidiospores. Descriptions, photographs and discussions are presented.

Keywords

Campeche, Macrofungi, Quintana Roo, tropical truffles, truffle-like fungi

Introduction

The species within *Stephanospora* Pat. were previously accommodated in Hymenogastraceae Vittad by Cunningham (1979) as *Octaviania* Vittad. and also in Octavianiaceae Locq. ex Pegler & T.W.K. Young by Pegler and Young (1979), due to the spiny basidiospores. They also included *Hydnangium* Wallr., *Sclerogaster* R. Hesse and *Wakefieldia* Corner &

Hawker. Nevertheless, Overwinkler and Horak (1979) placed these species in the family Stephanosporaceae Overwinkler & Horak along with *Lindtneria* Pilát. due to the spines around the basidiospore base, forming what is called a corona. Actually, Stephanosporaceae includes both sequestrate and resupinate species with or without a corona (Martín et al. 2004; Vidal 2004; Castellano et al. 2007; Lebel et al. 2015). *Stephanospora* is a genus with sequestrate species characterized by the subhypogeous habit, spiny or crested basidiospore ornamentation, and the conspicuous corona at the basidiospore base. Most of the species have a yellowish to orange pileus, pale-orange, olive-grey to pale-brown hymenophore and lack a stipe (Castellano et al. 1986; Pegler et al. 1993; Montecchi and Sarasini 2000; Vidal 2004). According to Lebel et al. (2015), 15 species are recognized worldwide.

Most Stephanospora species grow in association with broadleaf trees in Oceania (Cunningham 1979; Bougher and Lebel 2001; Lebel et al. 2015) or temperate forest in Europe (Palacios and Lakisbar 1991; Pegler et al. 1993; Vidal 2004; Fraiture and Novello 2013) and America (Vidal 2004; Guevara-Guerrero et al. 2015). Some additional undescribed species and genetic sequences were mentioned by Lebel et al. (2015) from Belize, Costa Rica and the Caribbean. In the USA, no species have been described from fruiting bodies, but DNA sequences have been included in a couple of analyses (Edwards and Zak 2010; Lebel et al. 2015). Most species can be found growing under mycorrhizal trees species such as *Podocarpus*, Eucalyptus, Quercus or Pinus, but no evidence of ectomycorrhizal associations has been observed (Tedersoo et al. 2010). The genus is represented in Mexico so far by a single species, S. michoacanensis Guevara & Castellano from central Mexico (Guevara-Guerrero et al. 2015).

In recent mycological exploration conducted by us on the Yucatán Peninsula in southern Mexico, some interesting sequestrate fungi were found, collected and identified as *Stephanospora*. The specimens were collected under *Haematoxylum campechianum* L., *Gymnopodium floribundum* Rolfe, *Metopium brownei* (Jacq.) Urband, and *Pinus caribaea* Morelet in lowland forest and pine savanna. Due to the basidiospore size, small corona, the association to lowland forest and pine savanna, and a molecular analyses of DNA we conclude that it is a novel species and we propose it as *S. mayana* de la Fuente, García-Jiménez, Guevara-Guerrero & Oros-Ortega.

Methods

Sampling data

Basidiomata were collected at Calakmul municipality in the state of Campeche and Othón P. Blanco municipality, in the state of Quintana Roo, Mexico. The vegetation is a disturbed lowland forest with *Coccoloba diversifolia* Jacq, *M. brownei*, *H. campechianum*, *G. floribundum*, and *Acoelorraphe wrightii* (Griseb. & H. Wendl.) H. Wendl. ex Becc. (Valdéz and Islebe 2011) and pine savanna with *P. caribaea*, *C. diversifolia*, *Curatella americana* L., *Crescentia cujete* L., and *Byrsonima crassifolia* (L.) Kunth (Macario and Sánchez 2011) (Fig. 1). Methods for collecting, sampling and



Figure 1. Habitat of Stephanospora mayana. a Lowland Forest at Blasillo b pine savanna at Xnohá.

describing sequestrate fungi were used (Castellano et al. 1986). Hand cuts sections were made from dried specimens mounted in KOH 5% and Meltzer reagent for microscopic description. Colour terminology was according to the Handbook of Colour (Kornerup and Wanscher 1978). All the specimens were curated and deposited at the mycological herbarium José Castillo Tovar of Instituto Tecnológico de Ciudad Victoria (ITCV).

Molecular analysis

For DNA extraction from basidiomata tissue we used the protocol reported by Cordova et al. (2014). Briefly, 0.1 g of the tissue was pulverized in liquid nitrogen and 1 ml extraction buffer of CTAB (20 mM EDTA pH 8.0, 100 mM Tris-HCl pH 8.0, 2% CTAB, 1.4 M NaCl, and 2-mercaptoethano) was add and incubated for 20–30 min at 65 °C and then vigorously mixed with a solution of phenol-chloroform isoamyl alcohol. After centrifugation, the supernatant was precipitated using cold isopropanol and sodium acetate and then incubated at –20 °C for 1 h. The DNA was pelleted by centrifugation and dried at room temperature. Finally, the DNA was resuspended in 100 µL of nuclease-free ultrapure water. Quantity and quality of the DNA was estimated with a NanoDrop ND-1000 spectrophotometer (NanoDrop Technologies, Wilmington, DE, USA).

The ITS region of the ribosomal DNA was amplified using the primers ITS1F/ITS4B reported by Gardes and Bruns (1993). The final concentration of the PCR reaction was: 1× of MyTaq reaction buffer, 0.4 µM of primer, 40 ng of DNA and 1.5 Unit of MyTaq DNA polymerase (Bioline, USA Inc.). The PCR conditions used for amplification were according to Gardes and Bruns (1993). The PCR products were observed on a 1.5 % agarose gel stained with ethidium bromide and visualized by UV transillumination in a Gel-DOC (Bio-Rad) equipment. Bands amplified were removed and purified with the QIA quick gel extraction kit (QIAGEN). Purified PCR products were sequenced using automated equipment in Davis Inc., CA, USA. Both sides of the cloned inserts were sequenced. Sequences were aligned with MUSCLE (Edgar 2004). Alignments were manually checked and ambiguous regions were excluded. Sequences produced in this study are deposited in GenBank under accession number MK033630. A search of GenBank nucleotide databank (NCBI) for homologous sequences was performed by BLAST analyses.

Phylogenetic analyses was performed from sequences obtained from basidiomata. References sequences (Lebel et al. 2015) and consensus sequence were aligned using BioEdit version 7.0.4.1 (Hall 1999). The tree was built in MEGA X (Kumar et al. 2018) using maximum likelihood analyses and the Kimura 2-parameter model (Kimura 1980) of nucleotide substitution with bootstrap values based on 1000 runs. *Piloderma fallax* and *Athelia arachnoidea* were used as outgroups (Lebel et al. 2015).

Results

Molecular analyses

A total of 48 sequences of *Stephanospora* species, including the new species, were analyzed (Fig. 2). The sequence consensus from the holotype clustered in the *Stephanospora* Clade III Subclade A (i) from Lebel et al. (2015). The designation of *S. mayana* as a new species is supported by ITS rDNA analyses and morphological features.



Figure 2. Phylogenetic tree inferred under the maximum-likelihood (ML) criterion from the ITS rDNA alignment corresponding the *Stephanospora* Clade III (i) dataset from Lebel et al. (2015). The tree was rooted using midpoint rooting. Numbers on the branches represent support values from 1,000 ML boostrap replicates. The branches are scaled in terms of the expected number of substitutions per site. Accession numbers in the sequence labels indicate sequences from GenBank.

Taxonomy

Stephanospora mayana de la Fuente, García-Jiménez, Guevara-Guerrero & Oros-Ortega, sp. nov.

MycoBank: MB 828118

Figure 3a-f

Holotype. Mexico: Campeche State, Calakmul Municipality, Blasillo town, 18°31'N, 88°18'W, 11 December 2017, de la Fuente (JF-397-ITCV), GenBank: MK033630.

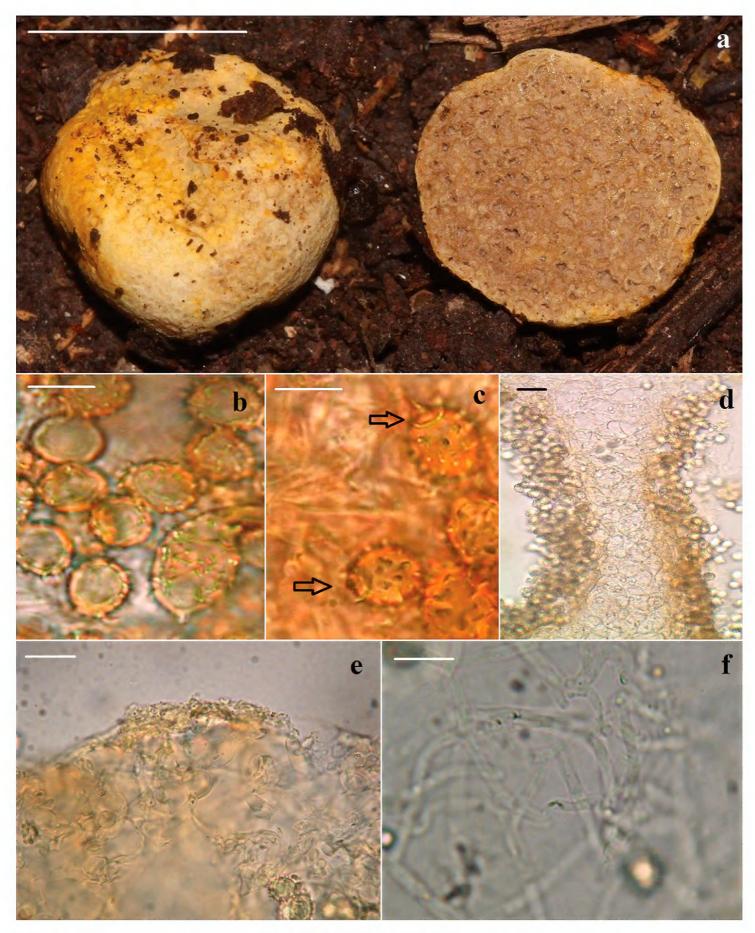


Figure 3. Stephanospora mayana (JF-397-ITCV-HOLOTYPE). **a** Basidiomata showing the pileus and hymenophore **b** basidiospores **c** corona **d** hymenophoral trama **e** pileus hyphae **f** hyphae from the locules. Scale bars: 10 mm (**a**); = $10 \mu \text{m}$ (**b, c, e, f**); $40 \mu \text{m}$ (**d**).

Diagnosis. Stephanospora mayana can be distinguished by the yellowish net-like pileus, the variable spore size $(8.0–17.0\times6.0–11.0~\mu m)$, thin pileus $(21.0–40.0~\mu m)$ and the ecological association to lowland forest and pine savanna with *H. campechianum*, *G. floribundum*, *C.diversifolia*, *M. brownei*, and *P. caribaea*.

Etymology. Named *mayana* in reference to the Mayan zone where this species was found.

Description. Basidiomata hypogeous to subhypogeous, scattered, $3.0-15 \times 2.0-6.0$ mm, globose to subglobose, without rizomorphs or stipe. Pileus yellowish to slightly orange (5A6-30A3-6), bruising pale orange when touched, wet to dry, sometimes net-like, dehiscent, showing locules inside. Hymenophore brittle, grayish (5C4), with empty rounded to angular locules, reaching 0.5 mm long, sometimes with white short and slender hyphae projecting from pileus to locules, trama sometimes orange (5A7-5B7), odour and taste strongly fruity.

Pileus 21.0–40.0 µm thick, composed of loosely interwoven, slender to inflated hyphae, 1.7–4.2 µm in diameter, orange to pale orange-yellow in KOH, thin-walled. Hymenophoral trama irregular, 62.0–100.0 µm wide, composed of irregular, globose, isodiametric and compacted hyphae, 13.5–26.3 µm in diameter, hyaline to slightly yellowish in KOH, thin-walled. Basidia 24.2–30.5 × 9.5–11.1 µm, clavate to subclavate, hyaline in KOH, guttulate, 2-spored, with long sterigmata, reaching 7 µm long, thin-walled, collapsing after basidiospore development. Basidiospores (8.0–) 10.0–16.0 (–17.0) × (6.0–) 8.5–10.5 (–11.0) µm (L = 12.10, W = 9.31, Q = 1.30, N = 90) ellipsoid to subglobose, with truncate to acute spines projecting 2.0 µm long, forming ridges reaching 3.5 µm high, sometimes coalescing, with a complete to partial corona 4.0–6.0 × 1.0–2.5 µm long, sometimes with 2–4 projecting spines, 1.5 µm long, with hilar appendage conspicuous, reaching 3 µm long, bright yellowish in KOH, orange in Meltzer reagent, with greenish to yellowish cell wall, 1.5–2.0 µm thick. Hyphae from the locules hyaline, 3.0–5.0 µm diameter, thin-walled. Clamp-connections absent in all tissues.

Distribution. Known from the Mexican states of Campeche and Quintana Roo where it is associated to lowland forest and pine savanna under *G. floribundum*, *H. campechianum*, *M. brownei*, and *P. caribaea*.

Additional material examined. Mexico, Quintana Roo, Othón Pompeyo Blanco municipality, Santa Elena Town, 18°30'N, 88°23'W, 07 October 2017, de la Fuente and Sánchez-Zavalegui 327 (Paratype); State of Campeche, Calakmul municipality, Xnohá town, 17°53'N, 89°10'W, 30 November 2017, de la Fuente 387 (paratype); Blasillo town, 18°31'N, 88°18'W, growing on abandoned termite mounds, 09 June 2018, de la Fuente 405 (paratype). (All in ITCV.)

Discussion

This species belongs to the *Stephanospora* clade III A (i) following Lebel et al. (2015). All the species in this clade are characterized by having basidiospores with ornamentation that does not project more than 2.5 μ m, basidia with sterigma up to 7 μ m long, and a small corona that never surpasses 7 μ m in width (Lebel et al. 2015). However, the Mexican material has larger basidiospores than any other species in this clade (up to 17 μ m), unlike *S. poropingao* T. Lebel & Castellano, *S. papua* T. Lebel & Castellano, *S. novae-caledoniae* T. Lebel, Castellano & K. Hosaka, and

Taxa	S. poropingao	S. novae-caledoniae	S. cribbae	S. рариа	S. mayana
Basidiomata size	5–25 mm	5–18 mm	5–25 mm	5–22 mm	3–13 mm
Pileus surface	Fibrillose	Smooth	Fibrillose	Irregular	Net-like
Pileus colour	Bright yellow, orangish-yellow to orangish-brown	Pale yellow to bright orange	Yellow to orange- yellow	Pale orange-yellow	Pale yellow
Hymenophore colour	Greyish-olive, olive- brown to yellow	White to pale yellow	Greyish olive to olive brown-yellow	Pale yellow	Cream, greyish olive
Odour	Not recorded	Faintly sweet	Faintly cocconut	Not recorded	Fruity
Basidiospores size	11–14 × 11–13 μm	11–14 × 09–12 μm	11–13.5 × 9.5–12 μm	09–11 × 07–8.5 μm	08–17 × 06–11 μm
Spines	Robust	Cylindrical, flattened to acute	Fine	Cylindrical or flattened	Truncated to acute
Corona size	05–09 × 01–03 μm	03–05 × 01–02 μm	03–05 × 01–02 μm	04–05 × 01–02 μm	04–06 × 01–2.5 μm
Pileus thickness	100–150 μm	40–145 μm	80–130 μm	30–140 μm	21–40 μm
Distribution	New Zealand, northwestern North Island	New Caledonia	Australia, Victoria, Queensland New South Wales	Papua New Guinea	Southern Yucatán Peninsula, Mexico
Habitat	Agathis-broadleaf, podocarp-broadleaf forest	Mixed forest with Nothofagus spp	Eucalyptus and Acacia Woodland	Mixed forest with Eucalyptus	Lowland forest and pine savanna

Table 1. Comparative morphology of *Stephanospora* species in the clade IIIA (i) according to Lebel et al. (2015).

S. cribbae T. Lebel & Castellano (up to 14 µm). Stephanospora kanuka T. Lebel & Castellano has similar pileus colour, sweet odour, and basidiospore length, but it has fine spines, an orange to yellowish hymenophore, and a fibrillose pileus (Lebel et al. 2015). Stephanospora cribbae T. Lebel & Castellano is similar to S. mayana in its yellowish pileus, corona size, and greyish hymenophore but differs in the smaller basidiospore size, the fibrillose pileus, and the coconut odour (Lebel et al. 2015). Stephanospora michoacanensis differs from S. mayana in having smaller basidiospores, the fruit odour absent, a cream colour pileus, and its association with oak-pine forest (Guevara-Guerrero et al. 2015). Stephanospora chilensis (E. Horak) J.M. Vidal differs in having an orange pileus and hymenophore, as well as smaller basidiospores (Vidal 2004).

Stephanospora mayana is in an unsupported clade with undescribed species from Belize (KM086881) and close to another unsupported clade with undescribed taxa from the USA and Spain (Lebel et al. 2015). Further collections and descriptions of taxa are required for Belizean and US material to better place this new species.

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References

- Bougher NL, Lebel T (2001) Sequestrate (truffle-like) fungi from Australia and New Zealand. Australian Systematic Botany 14: 439–484. https://doi.org/10.1071/SB00002
- Castellano MA, Trappe JM, Lodge DJ (2007) *Mayamontana coccolobae* (Basidiomycota), a new sequestrate taxon from Belize. Mycotaxon 100: 289–294.
- Castellano MA, Trappe JM, Maser Z, Maser S (1986) Key to Spores of the Genera of Hypogeous Fungi of North America, with Reference to Animal Mycophagy. Mad River Press, Eureka, 185 pp.
- Córdova I, Oropeza C, Puch-Hau C, Harrison N, Collí-Rodríguez A, Narvaez M, Nic-Matos G, Reyes C, Sáenz L (2014) A real-time PCR assay for detection of coconut lethal yellowing phytoplasmas of group 16SrIV subgroups a, D and E found in the Americas. Journal of Plant Pathology 96(2): 343–352.
- Cunningham GH (1979) The Gasteromycetes of Australia and New Zealand. J. Cramer, Vaduz, 236 pp.
- Dodge CW, Zeller SM (1936) *Hydnangium* and related genera. Annals of the Missouri Botanical Garden 23: 565–598. https://doi.org/10.2307/2394151
- Edgar RC (2004) Local homology recognition and distance measures in linear time using compressed amino acid alphabets. Nucleic Acids Research 32: 380–385. https://doi.org/10.1093/nar/gkh180
- Edwards IP, Zak DR (2010) Phylogenetic similarity and structure of Agaricomycotina communities across a forested landscape. Molecular Ecology 19: 1469–1482. https://doi.org/10.1111/j.1365-294X.2010.04566.x
- Fraiture A, Novello LA (2013) À propos de l'observation de *Stephanospora chilensis* en Italie. Scripta Botanica Belgica 51: 13–16.
- Gardes M, Bruns TD (1993) ITS primers with enhanced specificity for basidiomycetes-application to the identification of mycorrhizae and rusts. Molecular Ecology 2(2): 113–118. https://doi.org/10.1111/j.1365-294X.1993.tb00005.x
- Guevara-Guerrero G, Báez-Alvarado I, Gómez-Reyes VM, Castellano MA (2015) *Stephanos-pora michoacanensis* (Stephanosporaceae, Agaricales), a novel sequestrate truffle from North America. Revista Mexicana de Micología 41: 73–77.
- Hall TA (1999) BioEdit: a user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. Nucleic Acids Symposium Series 41: 95–98.
- Kimura M (1980) A simple method for estimating evolutionary rates of base substitutions through comparative studies of nucleotide sequences. Journal of Molecular Evolution 16: 111–120. https://doi.org/10.1007/BF01731581
- Kornerup A, Wanscher H (1978) Methuen Handbook of Colour. Eyre, London, 227 pp.
- Kumar S, Stecher G, Li M, Knyaz C, Tamura K (2018) MEGA X: Molecular Evolutionary Genetics Analysis across computing platforms. Molecular Biology and Evolution 35(6): 1547–1549. https://doi.org/10.1093/molbev/msy096
- Lebel T, Castellano MA, Beever RE (2015) Cryptic diversity in the sequestrate genus *Steph-anospora* (Stephanosporaceae: Agaricales) in Australasia. Fungal Biology 119: 221–218. https://doi.org/10.1016/j.funbio.2014.12.007

- Macario PA, Sánchez LC (2011) Pino tropical. In: Pozo C, Armijo N, Calmé S (Eds) Riqueza Biológica de Quintana Roo. Gobierno del Estado de Quintana Roo, Mexico City, 52–55.
- Martín MP, Raidl S, Tellería MT (2004) Molecular analyses confirm the relationship between *Stephanospora caroticolor* and *Lindtneria trachyspora*. Mycotaxon 90: 133–140.
- Montecchi A, Sarasini M (2000) Funghi Ipogei d'Europa. Associazione Micologica Bresadola, Fondazione Centro Studi Micologici, Vicenza, 714 pp.
- Overwinkler F, Horak E (1979) Eine neue Familie der Basidiomycetes mit aphylloporalen und gastroiden fruchtkörpern. Plant Systematics and Evolution 131: 157–164. https://doi.org/10.1007/BF00984129
- Palacios D, Lakisbar X (1991) *Stephanospora carotaecolor* (Berk.) Pat., una nueva cita para el Catálogo Micológico Ibérico. MUNIBE (Ciencias Naturales–NaturZientziak) 43: 115–119.
- Pegler DN, Spooner BM, Young TWK (1993) British Truffles. A Revision of British Hypogeous Fungi. The Board of Trustees of the Royal Botanical Gardens, Kew, 215 pp.
- Pegler DN, Young TWK (1979) The gasteroid Russulales. Translations from the British Mycological Society 72: 353–338. https://doi.org/10.1016/S0007-1536(79)80143-6
- Tedersoo L, May TW, Smith ME (2010) Ectomycorrhizal lifestyle in fungi: global diversity, distribution, and evolution of phylogenetic lineages. Mycorrhiza 20: 217–263. https://doi.org/10.1007/s00572-009-0274-x
- Valdéz M, Islebe G (2011) Tipos de vegetación en Quintana Roo. In: Pozo C, Armijo N, Calmé S (Eds) Riqueza Biológica de Quintana Roo. Gobierno del Estado de Quintana Roo, Mexico City, 32–75.
- Vidal JM (2004) The genus *Stephanospora* Pat., two new combinations. Revista Catalana de Micología 26: 97–111.